



# A new species of pterodectine feather mites (Acarina, Analgoidea, Proctophyllodidae) from the Little Spiderhunter Arachnothera longirostra (Passeriformes, Nectariniidae) in Meghalaya, India

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#### **Abstract**

The article describes a new species of the feather mite subfamily Pterodectinae from the Little Spider-hunter *Arachnothera longirostra* Temminck, 1826 (Passeriformes, Nectariniidae) in India. *Pedanodectes angustilobus* **sp. n.** differs from all known *Pedanodectes* species by having opisthosomal lobes short, at base wider than long, roughly rounded apically in males, and strongly elongated and narrowed lobar region with wide terminal appendages in females. A key to species of the genus *Pedanodectes* is presented.

## **Keywords**

Pterodectinae, Pedanodectes angustilobus, new species, systematics

## Introduction

Feather mites are commensals or ectoparasites permanently living on birds. In India, the diversity of feather mites is poorly investigated, and only 26 species have been mentioned so far in various taxonomic papers (Oudemans 1904, Bonnet 1924, Gaud and Mouchet 1963, Atyeo et al. 1972, Gaud 1972, McClure and Ratanaworabhan 1973, Gaud and Atyeo 1976, 1987, Santana 1976, Peterson et al. 1980, D'Souza and

Jagannath 1982, Atyeo 1984, Gaud et al. 1985, 1988, Dabert and Ehrnsberger 1998, Dabert 2003, Mironov et al. 2002, Putatunda et al. 2004, Constantinescu et al. 2014). Atyeo (in McClure and Ratanaworabhan 1973) reported five new species of feather mites collected in Asia from the host *Arachnothera longirostra* (including *Pedanodectes* species), but unfortunately this material has never been described.

In this paper, we describe a new species of *Pedanodectes* found on *A. longirostra* in India and we present a key for all species of this genus.

## Materials and methods

The material used in the present paper was collected in Meghalaya (India), in January 2014. The birds were trapped by means of ornithological mist nets, identified and visually checked for the presence of mites and, after mites were collected, released back to the wild. Mite specimens were placed in tubes with 95% ethanol. Later, in the laboratory, the mite specimens were cleared in lactic acid and mounted on microscope slides in Hoyer's medium. Drawings were made using an Olympus CX21 microscope, using a camera lucida drawing device.

The bird specimens were identified according to Rasmussen and Anderton (2012) and Grimmett et al. (2011), and the taxonomy of the birds used in the present paper follows Clements et al. (2013). The description of new species is given according to the current format used for species of pterodectine mites (Mironov and Fain 2003, Valim and Hernandes 2006, Mironov 2006, Mironov et al. 2008). The body chaetotaxy of mites follows that of Griffiths et al. (1990) with the modifications by Norton (1998) concerning coxal setae, and the chaetotaxy of legs follows Gaud and Atyeo (1996). The measuring techniques of particular structures used in the present paper were recently described by Mironov and Proctor (2009). We give the full set of measurements for the holotype (male) and a range of measurements for all corresponding paratypes. All measurements are in micrometers ( $\mu$ m). The holotype and all paratypes of the new species are deposited in the Acarological Collection of the "Grigore Antipa" National Museum of Natural History, Bucharest, Romania.

## Results

Family Proctophyllodidae Trouessart & Mégnin, 1884 Subfamily Pterodectinae Park & Atyeo, 1971

Genus Pedanodectes Park & Atyeo, 1971

The genus currently includes six species, associated with birds of the order Passeriformes (families Nectariniidae, Malaconotidae, Cisticolidae and Platysteiridae) in Africa (see Table 1).

Mite species	Host species	Host family	Location	References
P. hologaster (Gaud, 1953)	Chalcomitra senegalensis (Linnaeus)*, Chalcomitra fuliginosa (Bechstein)	Nectariniidae	Central African Republic	Gaud 1953; Park and Atyeo 1971
P. andrei (Till, 1954)	Tchagra senegalus (Linnaeus)	Malaconotidae	Mozambique	Till 1954
P. mesocaulus (Gaud & Mouchet, 1957)	Deleornis fraseri cameroonensis (Bannerman)	Nectariniidae	Cameroon	Gaud and Mouchet 1957
P. marginatus Mironov & Kopij, 1997	Camaroptera brachyura (Vieillot)	Cisticolidae	South Africa	Mironov and Kopij 1997
P. latior Mironov & Kopij, 1997	Platysteira peltata Sundevall	Platysteiridae	South Africa	Mironov and Kopij 1997
P. blaszaki Mironov, 2008	Cinnyris cupreus (Shaw)	Nectariniidae	South Africa	Mironov 2008
P. angustilobus sp. n.	Arachnothera longirostra (Latham)	Nectariniidae	India	Present paper

**Table 1.** Pedanodectes species and their host associations.

## Pedanodectes angustilobus Constantinescu, sp. n.

http://zoobank.org/186A8195-8BE0-4212-A864-0FB3A224DC66 Figs 1–4

**Type material.** Male holotype (ANA256), 3 male (ANA257, ANA258, ANA259) and 4 female (ANA260, ANA261, ANA262, ANA263) paratypes from the Little Spiderhunter *Arachnothera longirostra* Temminck, 1826 (Passeriformes, Nectariniidae); **IN-DIA:** Meghalaya, Jaintia Hills, Shnongrim village, (25°21'12.36"N, 92°31'3.06"E); 1151 m alt; 24.01.2014, collector Costică Adam.

**Description.** MALE (Figs 1A, B; 3A–D; holotype, range for 3 paratypes in parentheses): Length of idiosoma 316 (308-317), width 100 (100-104), length of hysterosoma 212 (207–212). Prodorsal shield divided into two parts by transversal band of soft tegument bearing setae se and si, antero-lateral extensions short and rounded, posterior margin slightly convex in median part, total length of shield 102 (102-106), greatest width 90 (84-90), surface without ornamentation (Fig. 1A). Scapular setae se separated by 38 (36–40). Humeral shields absent, setae cp situated ventrally, setae c2 situated dorsally, in anterior angles of hysteronotal shield. Subhumeral setae c3 lanceolate,  $20(19-20) \times 6(5-6)$ . Length of hysteronotal shield from anterior margin to bases of setae h2 195 (199-203), greatest width in anterior part 88 (80-86), anterior margin concave, anterior angles acute, surface without ornamentation with strongly sclerotized transverse fold between h1 setae. Opisthosomal lobes short, at base wider than long, roughly rounded apically, lobar apices bearing setae h2. Terminal cleft almost semicircular, with narrow membranous margin in anterior part, length of terminal cleft 10 (12–13). Supranal concavity absent. Hysteronotal setae c1, d1, e1 absent; setae h3 narrowly lanceolate, length 24 (24–25), greatest width 6 (4–6); setae h2 represented by macrosetae, length 130 (132–140), greatest width 5 (4–5); setae ps2 slightly thickened, 18 (12-16) long; setae ps1 filiform, minute, length 6 (5–6), situated slightly anterior to bases of setae h3 and h2, approximately equidistant

<sup>\* -</sup> Type host

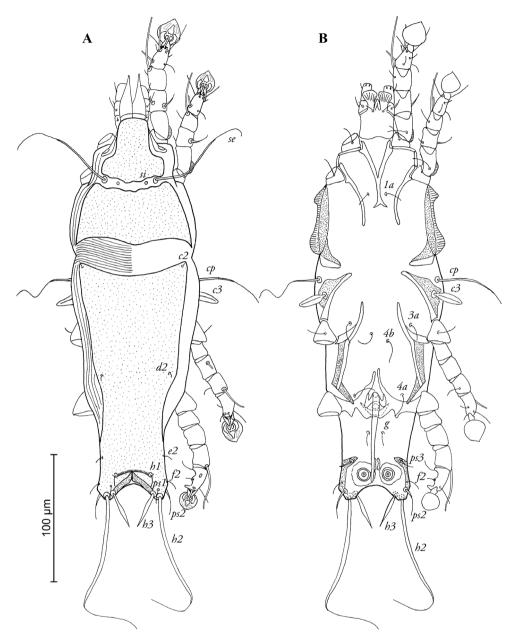
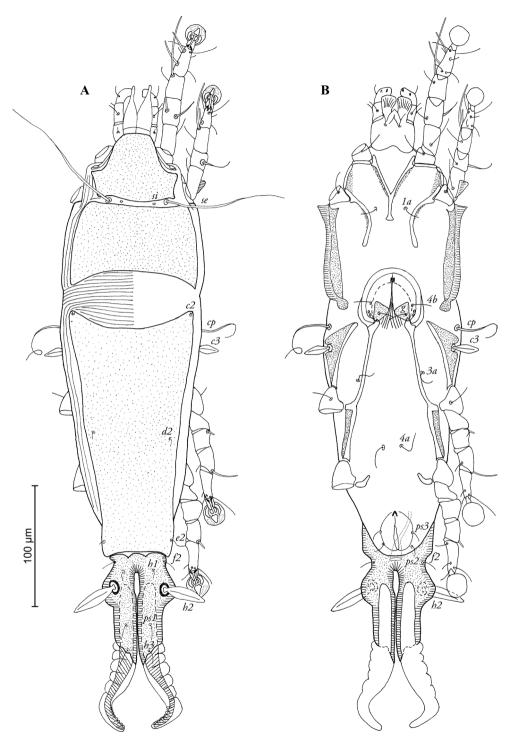


Figure 1. Pedanodectes angustilobus sp. n., male holotype: A dorsal view of idiosoma B ventral view of idiosoma.

from inner and outer margins of opisthosomal lobe. Dorsal measurements: *se-c2* 72 (66–70), *c2-d2* 92 (90–104), *d2-e2* 68 (60–66), *e2-h3* 32 (32–34), *h1-h3* 10 (10–12), *h2-h2* 38 (38–42), *h3-h3* 26 (26–28), *ps2-ps2* 50 (46–49). Epimerites I fused into a Y, sternum about ½ of the total length of epimerites, posterior end of sternum with pair

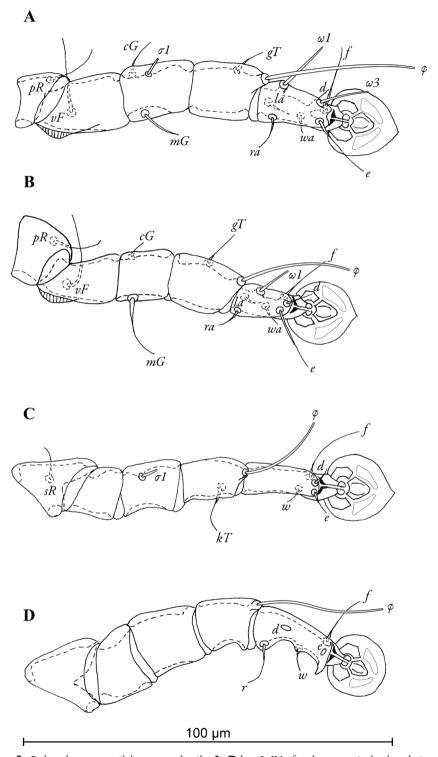


**Figure 2.** *Pedanodectes angustilobus* sp. n., female paratype: **A** dorsal view of idiosoma **B** ventral view of idiosoma.

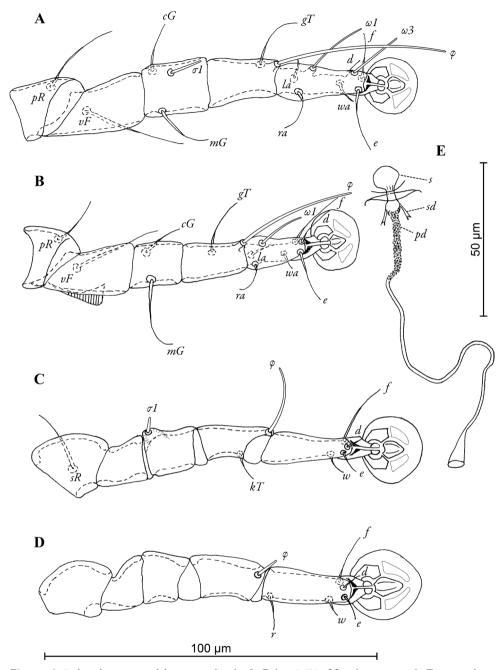
of postero-lateral extensions not connected to epimerites II. Epimerites II short, not extending to level of sejugal furrow. Coxal fields I–III open, without wide sclerotized areas. Epimerites IVa long, their anterior ends extending to midlevel of epimerites IV, their basal parts connected by semicircular transverse sclerite (supposedly genital shield) and almost completely encircling genital apparatus (Fig. 1B). Genital arch 11 (11–12) long, 14 (13–14) wide, basal sclerite of genital apparatus large, poorly sclerotized; aedeagus long, extending almost to anterior margin of terminal cleft, length of aedeagus from its anterior bend to tip 74 (72–78). Genital papillae indistinct, adanal shields absent. Anal suckers 11 (11–12) in diameter, corolla without indentations. Opisthoventral shields occupying lateral margin of opisthosoma, with narrow inner projection bearing seta *ps3*. Ventral measurements: *3a-4b* 22 (25–30), *4b-4a* 50 (44–50), *4a-g* 24 (24–28), *g-ps3* 32 (28–31), *ps3-ps3* 40 (32–38), *ps3-h3* 24 (23–24).

Legs I slightly longer and thicker than legs II, femora I and II with ventro-basal crests (Fig. 3A, B). Seta mGII strongly thickened in basal half. Tarsus IV 18 (19–22) long, with apical claw and with small apico-ventral extensions bearing seta r and w; setae d, e button-like, seta d bigger than e, situated in basal and apical parts of segment, respectively (Fig. 3D). Length of solenidia:  $\omega I$ I 15 (14–15),  $\omega I$ II 12 (13–16),  $\varphi$ I 42 (34–46),  $\varphi$ II 29 (32–34),  $\varphi$ III 22 (22–24),  $\varphi$ IV 32 (28–32).

FEMALE (Figs 2A, B; 4A-E; range for 4 paratypes): Length of idiosoma excluding terminal appendages 436-456, width 118-124, and length of hysterosoma 312-340. Prodorsal shield divided into two parts by transversal band of soft tegument, bearing setae se and si, antero-lateral extensions short and angular, posterior margin concave, total length of shield 118-122, greatest width 104-112, surface without ornamentation (Fig. 2A). Scapular setae se separated by 48-50. Humeral shields absent, setae cp situated ventrally, setae c2 situated dorsally, in anterior angles of hysteronotal shield. Subhumeral setae c3 lanceolate, 17–20 × 5–6. Hysteronotal shield divided into anterior hysteronotal shield and lobar shield. Anterior hysteronotal shield roughly rectangular, with anterior margin concave, greatest length 204-208, greatest width in anterior part 100-106, surface without ornamentation. Lobar shield elongated and narrowed, with well-developed lateral extensions bearing setae h2, length of lobar shield 86–92, width at level of setae h2 58–62. Terminal cleft narrow, almost parallelsided, with narrow elliptical part in anterior third, length 72-76, greatest width 3-4. Supranal concavity absent. Terminal appendages wide, their width in basal half similar to that of lobes. Hysteronotal setae c1, d1, e1 absent; setae h1 situated on lobar shield at level of anterior end of terminal cleft, setae h2 spindle-shaped, without terminal filaments, 38-40 × 6-9. Setae ps1 situated approximately equidistant from outer and inner margins of opisthosomal lobes, setae h3 6-8 long. Dorsal measurements: se-c2 88–94, *c2-d2* 100–112, *d2-e2* 86–94, *e2-h2* 36–44, *h2-h3* 48–52, *h1-h2* 16–22, *h1*h1 18-22, h2-h2 36-40, h3-h3 20-24. Epimerites I fused as a Y, posterior end of sternum with small rounded lateral extensions not reaching epimerites II. Coxal fields I, II open, without heavily sclerotized areas, outer margins of epimerites I and II with narrow sclerotized areas (Fig. 2B). Epimerites IVa rudimentary. Translobar apodemes of opisthosomal lobes present, fused to each other anterior to terminal cleft. Epigynum



**Figure 3.** *Pedanodectes angustilobus* sp. n., details: **A-D** legs I–IV of male, respectively, dorsal view.



**Figure 4.** *Pedanodectes angustilobus* sp. n., details: **A–D** legs I–IV of female, respectively **E** spermatheca and spermaducts, dorsal view. Abbreviations: pd - primary spermaduct; s - spermatheca; sd - secondary spermaduct.

horseshoe-shaped, outer margin with lateral extensions, greatest width 52–60. Head of spermatheca as in Fig. 4E, primary spermaduct with three different enlargements: ball-like enlargement near very head of spermatheca; moderate enlargement with verrucous external surface in proximal 1/5 of this duct; conical enlargement at copulatory opening. Secondary spermaducts short, not longer that ball-like enlargement of primary spermaduct. Copulatory opening situated ventral, posterior to anal opening. Distance between pseudanal setae: ps2-ps2 24–286, ps3-ps3 24–29, ps2-ps3 10–14.

Legs I longer and thicker than legs II, femora II with ventro-basal crests, genua III with dorso-basal crest (Fig. 4A–C). Genual setae mGI and mGII noticeably thickened in basal half. Length of solenidia:  $\omega II$  17–18,  $\omega III$  13–16,  $\varphi I$  45–56,  $\varphi II$  32–38,  $\varphi III$  22–27,  $\varphi IV$  7–8.

**Etymology.** The specific epithet refers to the narrowed lobar region of the female, and it is an adjective in the nominative singular.

Remarks. Pedanodectes angustilobus sp. n. clearly differs from all previously described Pedanodectes species because its males have well expressed opisthosomal lobes, and females have elongated and narrowed lobar region, with wide terminal appendages. Among previously known species of the genus, males of *P. angustilobus* sp. n. appear to be closest to that of *P. mesocaulus* (Gaud & Mouchet, 1957) from Deleornis fraseri cameroonensis (Bannerman, 1921) (Passeriformes: Nectariniidae). Males of both species have setae ps1 situated antero-lateral to the adamal suckers, epimerites I fused into Y, similar shape of opisthoventral shields and epimerites IVa with narrow anterior projections. Males of P. angustilobus sp. n. are easily to distinguish from those of *P. mesocaulus* by the following features: epimerites IVa have free the anterior projections and are connected posterior to the genital arch by a transversal sclerite, the tip of aedeagus does not extend beyond the posterior margin of idiosoma. In males of *P. mesocaulus*, the anterior projections of epimerites IVa are fused forming a pregenital sclerite while the transverse sclerite connecting the bases of these epimerites is absent, the tip of aedeagus extends beyond the posterior margin of idiosoma. Females of the new species are clearly different from those of the other species of the genus by the following unique combination of characters: the lobar region has the same width in anterior and posterior part; the terminal cleft is parallel-sided, with the margins almost touching, except for the anterior one third; and the terminal appendages are thick, their basal half is approximately as wide as opisthosomal lobes. In females of the other *Pedanodectes* species, the lobar region in the anterior part is wider than in the posterior part and the terminal appendages are narrower than lobes. The terminal cleft in females of the other species has the following shape: with the lateral margins parallel and almost touching in *P. andrei* and P. mesocaulus, with lateral margins sinuous and almost touching in certain parts in P. marginatus and P. latior; as a narrow inverted V in P. blaszaki, and as a narrow inverted U in P. hologaster.

## **Discussion**

According to the diagnosis of the genus *Pedanodectes*, some authors considered that males of this genus practically have no opisthosomal lobes and setae *ps3* are usually situated lateral or postero-lateral to the adanal suckers (Gaud and Atyeo 1996, Mironov 2008, Hernandes and Valim 2014). However in the original definition of this genus, Park and Atyeo (1971), apparently based on a some undescribed material they had on the hand, mentioned that the presence of weakly developed opisthosomal lobes and the position of setae *ps3* antero-lateral to the adanal suckers can be found in some species. The new species found on *A. longirostra* in India and described in this paper demonstrates the example of a *Pedanodectes* species having distinct opisthosomal lobes and setae *ps3* situated antero-lateral to the anal suckers in males.

# Key to males of Pedanodectes

1	Epimerites I fused V-likely2		
_	Epimerites I fused Y-likely4		
2	Postero-lateral extensions of epimerites I connected to epimerites II <i>P. latior</i>		
_	Postero-lateral extensions of epimerites I not connected to epimerites II3		
3	Tip of aedeagus extending beyond posterior margin of idiosoma <i>P. andrei</i>		
_	Tip of aedeagus not extending beyond posterior margin of idiosoma		
	P. hologaster		
4	Postero-lateral extensions of epimerites I present5		
_	Postero-lateral extensions of epimerites I absent		
5	Postero-lateral extensions of epimerites I connected to epimerites II		
	P. marginatus		
_	Postero-lateral extensions of epimerites I not connected to epimerites II6		
6	Opisthosomal lobes present; tip of aedeagus not extending beyond posterior		
	margin of idiosoma		
_	Opisthosomal lobes absent; tip of aedeagus extending beyond posterior mar-		
	gin of idiosoma		

# Key to females of Pedanodectes

1	Width of terminal appendages in anterior half similar to that of opisthosomal
	lobes
_	Terminal appendages narrower than opisthosomal lobes2
2	Lateral margins of terminal cleft spaced in some parts or on their entire
	length3
_	Lateral margins of terminal cleft almost touching on their entire length6

t level of scapular.	Prodorsal shield split into anterior and posterior pieces a	3
P. blaszaki	setae	
	Prodorsal shield entire	_
5	Lateral margins of prodorsal shield entire	4
und setae se	Lateral margins of prodorsal shield with deep incisions aro	_
P. hologaster	Epimerites I fused V-likely	5
P. marginatus	Epimerites I fused Y-likely	_
U	Posterior margin of anterior hysteronotal shield straight	6
	Posterior margin of anterior hysteronotal shield concave	_

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## References

- Atyeo WT (1984) A New Genus of Feather Mites and a New Expression of Male Polymorphism (Analgoidea: Avenzoariidae). Journal of the Kansas Entomological Society 57(3): 437–455.
- Atyeo WT, Gaud J, Humpfhreys WJ (1972) The feather mite genus *Freyanella* Dubinin, 1953 (Analgoidea: Pterollichidae). Acarologia 13: 383–409.
- Bonnet A (1924) Révision des genres *Megninia*, *Mesalges* et genres voisins de la sous-famille des sarcoptides plumicoles (2e Partie). Bulletin de la Société zoologique de France 49: 190–218.
- Clements JF, Schulenberg TS, Iliff MJ, Sullivan BL, Wood CL, Roberson D (2013) The eBird/Clements checklist of birds of the world. Version 6.8. http://www.birds.cornell.edu/clementschecklist/download/ [accessed 20 January 2014]
- Constantinescu IC, Chişamera G, Mukhim KB, Adam C (2014) Two new feather mite species of the family Pteronyssidae (Acarina: Analgoidea) from Meghalaya (Northeast India). Zootaxa 3774(4): 351–366. doi: 10.11646/zootaxa.3774.4.4
- Dabert J, Ehrnsberger R (1998) Phylogeny of the feather mite family Ptiloxenidae Gaud, 1982 (Acari: Pterolichoidea). Biosystematics and Ecology Series 14: 145–178. http://www.landesmuseum.at/pdf\_frei\_remote/BioEco\_14\_0145-0178.pdf
- Dabert J (2003) The feather mite family Syringobiidae Trouessart, 1896 (Acari, Astigmata, Pterolichoidea). I. Systematics of the family and description of new taxa. Acta Parasitologica 48: S1–S184.

- D'Souza PE, Jagannath MS (1982) A new genus and species of syringobiid mite (Acari) in the quill of domestic fowl (*Gallus domesticus*) from south India. Indian Journal of Acarology 6: 51–56.
- Gaud J (1953) Sarcoptidés plumicoles des oiseaux d'Afrique occidentale et centrale. Annales de parasitologie humaine et comparée 28: 193–226.
- Gaud J (1972) Acariens Sarcoptiformes plumicoles (Analgoidea) parasites sur les oiseaux Charadriiformes d'Afrique. Musée Royal de l'Afrique Centrale, Annales Sciences Zoologiques 193: 1–116.
- Gaud J, Atyeo WT (1976) Discordances entre les aires de répartition géographique des parasites et celles de leurs hotes chex les sarcoptiformes plumicoles. Acarologia 18: 329–344.
- Gaud J, Atyeo WT (1987) Les *Trouessartia* (Analgoidea, Trouessartiidae) parasites des hirondelles de l'ancien monde II. Le groupe Minutipes. Acarologia 28: 367–379.
- Gaud J, Atyeo WT (1996) Feather mites of the world (Acarina, Astigmata): the supraspecific taxa. Musée Royal de l'Afrique Centrale, Annales Sciences Zoologiques 277: 1–191.
- Gaud J, Mouchet J (1957) Acariens plumicoles (Analgesoidea) des oiseaux du Cameroun. I. Proctophyllodidae. Annales de parasitologie humaine et comparée 32: 491–546.
- Gaud J, Mouchet J (1963) Révision des genres *Grallobia* Hull et *Grallolichus* Gaud (Pterolichidae, Sarcoptiformes). Acarologia 5: 628–643.
- Gaud J, Atyeo WT, Barré N (1985) Les acariens du genre *Megninia* (Analgidae) parasites de *Gallus gallus*. Acarologia 26(2): 171–182.
- Gaud J, Rosen S, Hadani A (1988) Les Acariens plumicoles du genre *Megninia* parasites des poulets domestiques. Science véterinaires médicine comparée 90: 83–98.
- Griffiths DA, Atyeo WT, Norton RA, Lynch CA (1990) The idiosomal chaetotaxy of astigmatid mites. Journal of Zoology 220: 1–32. doi: 10.1111/j.1469-7998.1990.tb04291.x
- Grimmett R, Inskipp C, Inskipp T (2011) Helm Field Guides: Birds of the Indian Subcontinent. Christopher Helm, London, 528 pp.
- Hernandes FA, Valim MP (2014) On the identity of two species of Proctophyllodidae (Acari: Astigmata: Analgoidea) described by Herbert F. Berla in Brazil, with a description of *Lamellodectes* gen. nov. and a new species. Zootaxa 3794(1): 179–200. doi: 10.11646/zootaxa.3794.1.8
- McClure HE, Ratanaworabhan N (1973) Some ectoparasites of the birds of Asia. Jintana Printing Ldt, Bangkok, 219 pp.
- Mironov SV (2006) Feather mites of the genus *Montesauria* Oudemans (Astigmata: Proctophyllodidae) associated with starlings (Passeriformes: Sturnidae) in the Indo-Malayan region, with notes on systematic of the genus. Acarina 14: 21–40.
- Mironov SV (2008) Three new species of the feather mite subfamily Pterodectinae (Acari: Astigmata: Proctophyllodidae) from passerines (Aves: Passeriformes) in Central Africa. Annales Zoologici 58 (2): 403–418. doi: 10.3161/000345408X326744
- Mironov SV, Fain A (2003) New species of feather mite subfamily Pterodectinae (Astigmata: Proctophyllodidae) from African passerines (Aves: Passeriformes). Bulletin de la Société Royale Belge d'Entomologie 139: 75–91.
- Mironov SV, Kopij G (1997) New feather mite species of the subfamily Pterodectinae (Astigmata: Proctophyllodidae) from some passerines (Aves: Passeriformes) of South Africa. Journal of African Zoology 111: 449–463.

- Mironov SV, Proctor HC (2009) Feather mites of the genus *Proterothrix* Gaud (Astigmata: Proctophyllodidae) from parrotbills (Passeriformes: Paradoxornithidae) in China. Journal of Parasitology 95: 1093–1107. doi: 10.1645/GE-1961.1
- Mironov SV, Dabert J, Ehrnsberger R (2002) On systematics of the feather mite genus *Triphyllochaeta* Dubinin, 1956 (Astigmata: Pterolichidae). Entomologische Mitteilungen aus dem Zoologischen Museum Hamburg 14: 27–37.
- Mironov SV, Literak I, Čapek M (2008) New feather mites of the subfamily Pterodectinae (Acari: Astigmata: Proctophyllodidae) from passerines (Aves: Passeriformes) in Mato Grosso do Sul, Brazil. Zootaxa 1947: 1–38.
- Norton AR (1998) Morphological evidence for the evolutionary origin of Astigmata (Acari: Acariformes). Experimental & Applied Acarology 22: 559–594. doi: 10.1023/A:1006135509248
- Oudemans AC (1904) Acarologische Aanteekeningen XIV. Entomologische Berichten 1: 190–195.
- Peterson PC, Atyeo WT, Moss WW (1980) The feather mite family Eustathiidae (Acarina: Sarcoptiformes). Monograph. Academy of Natural Sciences of Philadelphia 21: 1–143.
- Park KC, Atyeo WT (1971) A generic revision of the Pterodectinae, a new subfamily of feather mites (Sarcoptiformes: Analgoidea). Bulletin of the University of Nebraska State Museum 9(3): 40–88.
- Putatunda BN, Kumar R, Banerjee DP (2004) Mites parasitic and/or associated with poultry and some mammals in Hisar, Haryana with key to identification. Indian Journal of Animal Research 38: 1–7.
- Rasmussen PC, Anderton JC (2012) Birds of South Asia. The Ripley Guide. Volumes 1 and 2. Second Edition. National Museum of Natural History Smithsonian Institution, Michigan State University and Lynx Edicions, Washington, D.C., Michigan and Barcelona, vol. 1: 684 pp., vol. 2: 378 pp.
- Santana FJ (1976) A review of the genus *Trouessartia* (Analgoidea: Alloptidae). Supplements to the Journal of Medical Entomology 1: 1–128.
- Till WM (1954) Five new feather mites of the genus *Pterodectes* (Acarina: Analgesoidea). Moçambique 79: 85–100.
- Valim MP, Hernandes FA (2006) Redescription of four species of the feather mite genus *Pterodectes* Robin, 1877 (Acari: Proctophyllodidae: Pterodectinae) described by Herbert F. Berla. Acarina 14: 41–55.